Research Paper

# Theobromine is associated with slower epigenetic ageing

Ramy Saad<sup>1,2</sup>, Ricardo Costeira<sup>1</sup>, Pamela R. Matías-García<sup>3</sup>, Sergio Villicaña<sup>1</sup>, Christian Gieger<sup>3</sup>, Karsten Suhre<sup>4</sup>, Annette Peters<sup>5,6</sup>, Gabi Kastenmüller<sup>7</sup>, Ana Rodriguez-Mateos<sup>8</sup>, Cristina Dias<sup>9</sup>, Cristina Menni<sup>1,10,11</sup>, Melanie Waldenberger<sup>3</sup>, Jordana T. Bell<sup>1</sup>

<sup>1</sup>Department of Twin Research and Genetic Epidemiology, King's College London, St Thomas' Hospital Campus, London SE1 7EH, UK

<sup>2</sup>North East Thames Regional Genetic Service, Great Ormond Street Hospital for Children NHS Foundation Trust, London, UK

<sup>3</sup>Research Unit Molecular Epidemiology, Institute of Epidemiology, Helmholtz Zentrum München, German Research Center for Environmental Health, Neuherberg, Germany

<sup>4</sup>Department of Physiology and Biophysics, Weill Cornell Medicine - Qatar, Education City - Qatar Foundation, Doha, Qatar

<sup>5</sup>Institute of Epidemiology, Helmholtz Zentrum München, German Research Center for Environmental Health, Neuherberg, Germany

<sup>6</sup>Institute for Medical Informatics, Biometrics and Epidemiology, Ludwig-Maximilians-Universität (LMU) Munich, Munich, Germany

<sup>7</sup>Institute of Computational Biology, Helmholtz Zentrum München, German Research Center for Environmental Health, Neuherberg, Germany

<sup>8</sup>Department of Nutrition, King's College London, St Thomas' Hospital Campus, London SE1 7EH, UK

<sup>9</sup>Department of Medical and Molecular Genetics, School of Basic and Medical Biosciences, Faculty of Life Sciences and Medicine, King's College London, London, UK

<sup>10</sup>Department of Pathophysiology and Transplantation, Università Degli Studi di Milano, Via Francesco Sforza, Milan 20122, Italy

<sup>11</sup>Fondazione IRCCS Cà Granda Ospedale Maggiore Policlinico, Angelo Bianchi Bonomi Hemophilia and Thrombosis Center, Milan 20122, Italy

Correspondence to: Ramy Saad, Jordana T. Bell; email: ramy.saad@kcl.ac.uk, jordana.bell@kcl.ac.uk

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### **ABSTRACT**

Theobromine, a commonly consumed dietary alkaloid derived from cocoa, has been linked to extended lifespan in model organisms and to health benefits in humans. We examined associations between circulating levels of theobromine intake, measured using serum metabolomics, and blood-based epigenetic markers of biological ageing in two European human population-based cohorts. Serum theobromine levels were significantly associated with reduced epigenetic age acceleration, as measured by GrimAge (p < 2e-7) and DNAmTL (p < 0.001) in 509 individuals from the TwinsUK cohort, and both signals replicated in 1,160 individuals from the KORA cohort (p = 7.2e-08 and p = 0.007, respectively). Sensitivity analyses including covariates of other cocoa and coffee metabolites suggest that the effect is specific to theobromine. Our findings indicate that the reported beneficial links between theobromine intake on health and ageing extend to the molecular epigenetic level in humans.

### **INTRODUCTION**

Dietary phytochemicals are compounds found in plants that have been reported to benefit human health. They include polyphenols, alkaloids, terpines, flavonoids, and others [1]. Evidence from both epidemiological and human intervention trials have identified beneficial effects of various phytochemicals on health and ageing, including on biomarkers of cholesterol transport [2], inflammation [3], and cellular senescence [4].

Alkaloids in plants form a large component of dietary phytochemicals, as they are both abundant and highly bioactive [5]. This bioactivity is a function of their purpose as protective chemicals and, therefore, alkaloids have wide-ranging *in vivo* actions, along with narrow therapeutic indices. Specifically, alkaloids have been studied for their relevance to age-related diseases, including cancers [6], type 2 diabetes [7] and inflammation [8]. Notable examples of pharmacologically active alkaloids include indole [9], indolizidine [10], as well as specific subtypes such as berberine [11], morphine, strychnine, quinine, and others [5].

Coffee and cocoa are widely consumed foods, associated with reduced cardiovascular disease (CVD) and mortality [12, 13]. Cocoa and coffee share several important alkaloids including the methylxanthines theobromine (TB), caffeine (CAF), theophylline (TP), paraxanthine (PX) and 7-methylxanthine (MX) [14] (Figure 1A). The coffee-associated methylxanthines (CAF, TP and PX) are found in lower concentrations in cocoa [8]. TB and MX, are partial metabolites of CAF, though both are also found in much higher concentrations in cocoa as primary unprocessed metabolites [15]. TB has previously been linked to multiple aspects of health and ageing. For example, studies in model organisms have identified links between TB and extended lifespan [16]. Furthermore, multiple observational human cohort studies have reported clear links between TB intake and various aspects of improved health [17]. Despite this, the exact impacts of TB on health and ageing are still not fully understood, and the molecular pathways that underlie these effects are largely unknown.

Multiple biological mechanisms can mediate the effects of dietary phytochemicals on human health and

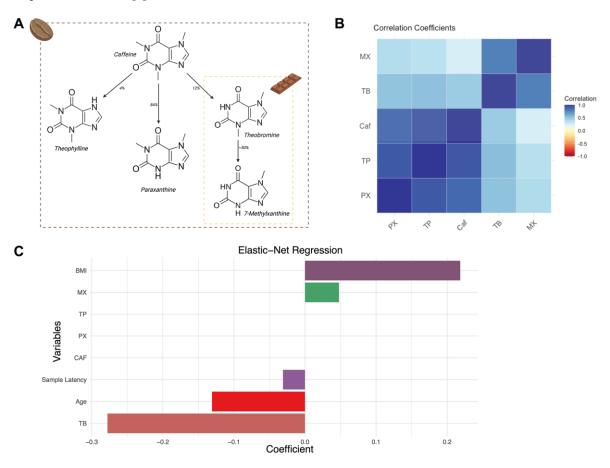


Figure 1. Major dietary sources of methylxanthines and their correlations in the TwinsUK sample. (A) Schematic presenting key methylxanthines, their respective dietary sources and their derivation as secondary metabolites. (B) Correlation heatmap of coffee-related metabolites in the TwinsUK sample. (C) Bar plot representation of the Elastic-net Regression coefficients with 10-fold cross-validation of variables against GrimAgeAccel in the TwinsUK sample.

ageing, and one of these is the epigenetic regulation of gene expression. Alkaloids can influence epigenetic processes, for example, through inhibition of histone deacetylases or DNA methyltransferases (DNMTs) [18]. Cocoa and coffee consumption have been linked to multiple DNA methylation changes in humans, where extracts from cocoa can affect global leukocyte DNA methylation levels potentially though inhibition of DNMTs [19], and distinct blood DNA methylation signals have been associated with coffee consumption [20]. Therefore, alkaloids, such as those found in cocoa, may exert their beneficial effects on health and ageing potentially through changing the human epigenome.

Epigenetic deregulation is a key hallmark of ageing. The effects of ageing on genome-wide methylation have been widely documented, including reduction of global DNA methylation [21], global increase in Shannon entropy of methylation patterns [22], and site-specific changes in differential and variable DNA methylation levels [23-25]. Hence, multiple studies have developed epigenetic clocks towards predicting different agerelated features, such as chronological age [26], time to death [27], pace of ageing [28], as well as other molecular biomarkers of ageing including telomere length [29]. As such, epigenetic clocks may act as useful tools for assessing whether specific dietary phytochemicals are associated not only with epigenetic modifications, but also with the rate of ageing, as measured by these clocks.

Several recent studies have investigated the association of nutrients and metabolites to epigenetic ageing. Some studies have focused on diet questionnaire data, identifying associations between vitamins B and C intakes with epigenetic ageing [30]. Smaller-scale intervention trials have also explored the impact of dietary changes on epigenetic age. For example, an 8week randomised controlled trial intervention in six post-menopausal women found that an increase in dietary polyphenols resulted in significant deceleration of epigenetic aging, as measured by the Horvath clock [31]. Moreover, dietary interventions such as calorie restriction can also influence epigenetic aging, as identified from the CALERIE trial using the DunedinPACE epigenetic clock [32]. However, results were not consistent across different epigenetic clocks, highlighting potential variability in how they capture ageing processes.

In this study, including two independent human population-based cohorts, we investigated whether individual bioactive alkaloids in coffee and cocoa are associated with reduced epigenetic ageing, and may therefore potentially contribute towards extension of human healthspan.

#### **RESULTS**

We initially tested for the association between six metabolites found in coffee and cocoa, and epigenetic measures of ageing in blood samples from 509 healthy females from the TwinsUK cohort (median age = 59.8, IQR = 12.81, BMI = 25.35). The six metabolites included the methylxanthines CAF, TP, TB, MX and PX and theanine, and biological ageing analyses focused on GrimAgeAccel. TB was significantly associated with reduced epigenetic ageing as captured by GrimAgeAccel (B = -1.576, standard error = 0.3, p = 3.99e-6) (Figure 2, Supplementary Table 1). This was significant at Bonferroni Correction (p < 0.0083). Extending analyses to test for association between TB and three other measures of biological ageing, including methylation markers of telomere length, PhenoAge and DunedinPACE, we identified another significant association with DNAmTL (B = 0.03, standard error = 0.0124, p = 0.0029) (Figure 2, Supplementary Table 1). This was significant at Bonferroni correction (p < 0.0125).

As both cocoa and coffee include TB, we carried out further analyses to dissect the correlations between methylxanthines and food component intakes within the TwinsUK sample. Correlation coefficients support the expected patterns of close correlation among coffee-associated methylxanthines (CAF, TP, PX) and close correlation among the cocoa-associated methylxanthines (TB and MX) (Figure 1B), demonstrating consistency with the undertaken metabolomic analysis.

Specifically, we observed that coffee-associated methylxanthines CAF and TP were strongly correlated to each other (R = 0.89), and that cocoa-associated methylxanthines TB and MX were also strongly correlated to each other (R = 0.78). In contrast, TB and CAF showed only moderate correlation (R = 0.46) (Figure 1B). The weaker correlation between TB and CAF in our cohort reflects the expected low metabolism of CAF to TB, and the likely differential food sources of these metabolites in vivo [33]. Indeed, TB was previously associated with chocolate consumption in a larger sample from the same TwinsUK population cohort (B = 0.024, p = 1.34e-11) [34]. In the current sample (509 twins), we confirm a positive, but weaker correlation between the consumption of 'chocolate' (as reported by food frequency questionnaires) and TB levels (R = 0.136). TB consumption was not strongly associated to diet quality (AHEI, R = -0.0293).

Several sensitivity analyses were undertaken to mitigate the effects of potential dietary intake confounders. First, we re-examined the association between TB and GrimAge acceleration, but now including CAF, TP, PX, and MX as additional covariates. We then carried out similar analyses, also including TP, PX, and MX as additional covariates because they are metabolite derivatives of CAF [14] (Figure 1A). The associations between TB and slower epigenetic ageing remained significant in these extended models with additional covariates (n = 509, B = -0.823, SE = 0.268, p = 0.00219; (Supplementary Figure 1A, 1B;

Supplementary Table 1), suggesting that effects are specific to TB and not an alternative xanthine derivative. Furthermore, we re-analysed the data to assess the effect of time latency between date of DNA methylation and metabolomic sample collection. Samples were subset by window of latency periods, including 2 years (n = 420, B = -0.724, p = 1.03e-5), 1 year (n = 276, B = -0.75, p = 0.00015) and

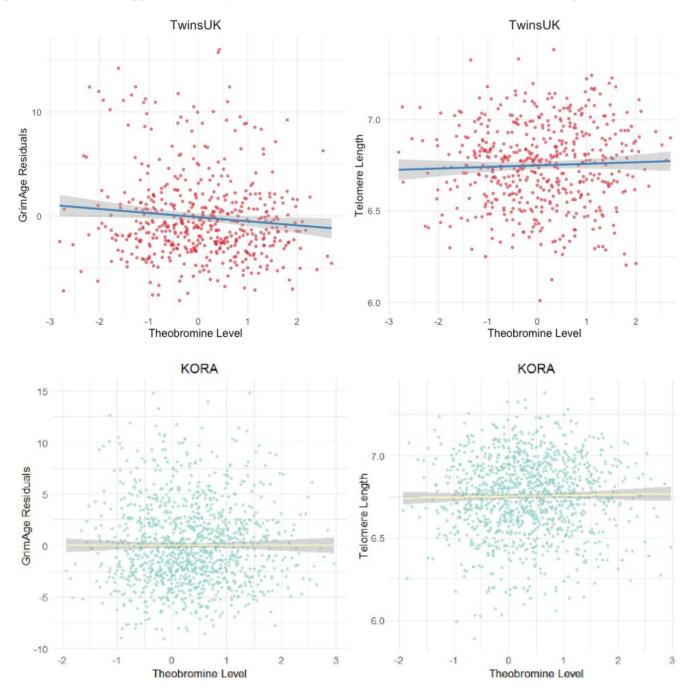


Figure 2. The association between TB and epigenetic age in the TwinsUK and KORA cohort samples. Scatter plots of the GrimAge acceleration residuals (top left, primary analysis B = -1.576,  $p = 3.99 \times 10^{-6}$ ) and epigenetic estimate of telomere length, DNAmTL (top right, primary analysis B = 0.036, p = 0.003), in the discovery TwinsUK sample of 509 females. Bottom row plots show results for the KORA sample of 1,160 individuals for GrimAge acceleration residuals (bottom left, primary analysis B = -1.06,  $p = 7.2 \times 10^{-6}$ ) and DNAmTL (bottom right, primary analysis B = 0.022, p = 0.007).

contemporaneous (same-day) sampling (n = 121, B = -1.576, p = 3.99e-6). The strength of the association increased with shorter latency periods (Supplementary Figure 1C; Supplementary Table 1).

Targeted replication of the TB and epigenetic ageing rate associations was sought in a larger sample of 1,160 individuals from the KORA cohort [35], where serum metabolomic and DNA methylation profiles were obtained from the same time-point. We replicated the association between reduced GrimAgeAccel and TB in a model including all technical and biological covariates of our study (CAF, TP, PX, MX; coefficient = -1.06, standard error = 0.195, p = 7.177E-08) (Figure 2). We also observed a significant association with GrimAge in the subset of females alone in the KORA sample (n = 592, B = -0.79, p = 0.0022). We also replicated the association of TB with DNAmTL (coefficient = 0.022, standard error = 0.008, p = 0.007) (Figure 2).

As a further follow-up analysis, we also stratified the TwinsUK sample by smoking status. The reduced epigenetic ageing acceleration signal was most significant in previous and current smokers (B = -2.687, p = <2.2e-16, n = 53), compared to never smokers (Supplementary Figure 1A, 1B). Nicotine likely induces the breakdown of TB by enzyme induction [36] and may influence the pharmacodynamic clearance or bioavailability of TB and its byproducts.

A final set of follow-up analyses explored feature selection using LASSO and elastic net regression to assess which metabolites most strongly relate to the epigenetic measures of biological ageing. LASSO Regression is a penalisation technique, using shrinkage, to reduce the impact of variables with high collinearity. LASSO regression with GrimAgeAccel as the dependent variable and all technical covariates and metabolites as independent variables identified TB as a significant predictor for GrimAgeAccel (coefficient = -0.231; RMSE = 3.644), with similar results using 10fold cross-validation (coefficient = -0.186; RMSE = 3.834). We utilised Elastic-net regression, to adjust for potential over-penalisation of collinearity that can occur with LASSO (Elastic-net regression utilises a spectrum of penalisation between absolute sums, LASSO, alpha = 0.1, to squared sums, ridge penalisation, alpha = 1). Elastic-net regression with 10- fold cross-validation (best alpha: 0.2, best lambda: 0.419) showed consistent results (GrimAgeAccel TB coefficient = -0.277; RMSE = 3.8) (Figure 1C).

### **DISCUSSION**

Here we report a significant association between circulating levels of theobromine (TB) with slower

epigenetic ageing in two independent population-based cohorts. TB is a relatively unexplored dietary phytonutrient that has recently been linked to beneficial health effects and extended lifespan in model organisms [16]. However, there have been limited studies of the role of TB in human cohorts.

The association of TB and biological ageing measures is most pronounced by the GrimAge epigenetic clock acceleration measures, which strongly predicts time to death. The pattern was also captured by DNAmTL, which estimates telomere length. The two epigenetic ageing measures, DNAmTL and GrimAge acceleration residuals are weakly correlated (R = 0.29) in the TwinsUK sample, and this supports previous reports in the literature that telomere length and genome-wide epigenetic ageing are independently associated with ageing [37]. Previous work has explored how epigenetic clocks may capture different mechanisms underpinning hallmarks of ageing, such as telomere attrition and epigenetic ageing [38]. We therefore considered the two measures to capture separate aspects of the ageing process, that do not necessarily overlap.

Methylxanthines are found across various food groups in different proportions, with CAF being the most prominent in coffee, and TB being the most prominent in cocoa [39]. Exact proportions can vary across foods and also depend on food quality, processing methods (such as decaffeination), or inter-individual variability (such as genetic variation in monooxygenase function or presence of exogenous P450 enzyme inducers or inhibitors [40]). Our sensitivity analyses support the conclusion that the association effect is specific to TB and is likely not attributed to CAF, TP, PX or MX. This conclusion stems from results based on accounting for multiple metabolites as covariates in the linear association models, and results from 10-fold crossvalidation LASSO and elastic-net regression. This suggests that TB may affect a common biological pathway relevant to ageing.

Several studies predominantly in model organisms have identified links between TB and improved aspects of health and ageing. Importantly, TB has been reported to extend lifespan in ROS-sensitive strains of *C. elegans* [16]. It has also been noted to have differential psychotropic actions to caffeine [41]. In mice, modest supplementation of 0.05% TB results in significant increases in the neurotrophic factors CREB and BDNF, which are relevant to reward and learning [42], but higher doses of TB were associated with better lipid profiles and lower blood pressure in a retrospective cross-sectional study [43]. Although some methyl-xanthines are used in clinical practice [44], TB has not been explored in depth for its medical utility, but it has

been suspected to be of importance to human health [45]. TB has also been previously associated with the enrichment of beneficial microbiota with SFCA-producing abilities [46]. Future studies should explore if the gut microbiome composition may mediate the effect of TB on human health and ageing.

While we found replication of our signals across both cohorts, it is interesting to note the differences between them. The female-specific TwinsUK cohort replication with the mixed KORA cohort suggests that a sexspecific effect is not a key factor in the association. Indeed, when the KORA cohort was subsequently subsetted to only females, we found a reduction in effect size for GrimAge (n = 592, B = -0.79, p = 0.0022); suggesting that the results are not sex-specific. Future replication in diverse cohorts is warranted to investigate any cohort-specific effects further.

The differential effects observed for smoking status are also compelling and suggest that the effects of TB could be more pronounced in smokers. It is of note that, in addition to smoking status being a key variable in the development of GrimAge, many of the differential DNA methylation changes associated with smoking can be responsive to cessation [47]. Further research, including experimental work, is needed to confirm and dissect further these differential effects.

One important limitation in the discovery cohort is the latency between metabolomic and epigenetic sample acquisition, which may be a source of bias. Our latency-stratified analysis and the contemporaneous samples acquired in the Replication cohort, however, suggest that latency-associated bias is not a key confounder of the observed association.

One possible explanation for the correlation between TB and epigenetic age is whether it may be a biomarker for a collinear confounder. For instance, TB may signify flavan-3-ol consumption, as these (poly)phenols are abundant in cocoa but were not available in the metabolomic data. Methylxanthines, including theobromine, have been shown to enhance the vascular effects of flavan-3-ols by improving endothelial function and increasing nitric oxide bioavailability; however, when administered alone, they did not elicit any effect [48] and the cardiometabolic and healthy aging benefits of flavan-3-ols are well established [13, 49].

On the other hand, the sensitivity analysis using elasticnet regression supports the conclusion that the effect is specific to TB and no other collinear methylxanthines, making the possibility of a hidden confounding variable less likely. Further research is needed to disentangle the potential mechanisms by which TB is associated with reduced epigenetic ageing and exclude any potential confounders not assessed by our study or reverse-causality.

In conclusion, our study identifies an association between TB and measures of epigenetic ageing, suggesting that TB is relevant to human ageing. Further exploration of TB and age-related health markers may identify key epigenetic mechanisms transducing this effect and reveal a potential use of TB towards extending the human healthspan.

## MATERIALS AND METHODS

## Discovery cohort data

The discovery sample in this study included 509 monozygotic and dizygotic twin female participants from the TwinsUK cohort [50]. This constituted the total number of samples with relevant data acquired from the TwinsUK cohort (14,838 twins). Median age was 58.9 years with a standard deviation of 8.79 years, showing an approximately Normal distribution with a slight right skew (Shapiro Wilk Normality = 0.993, p = 0.0283, adjusted Fisher-Pearson skew = +0.091). Altogether, 228 samples were from never-smokers.

Metabolomic data in these participants were generated in fasting serum samples using the Metabolon Inc mass spectrometry platform (Metabolon, Inc., Durham, NC). Metabolite concentrations were measured at fasting from serum, samples by Metabolon Inc. (Durham, USA) using an untargeted Liquid chromatographymass spectrometry (LC-MS) platform as previously described [51]. Metabolites with more than 20% missingness and metabolite data outliers (±3SD from mean) were excluded. The remaining metabolites were day median-normalised, imputed to the day minimum, and inverse-normalised. Six metabolites associated with coffee or cocoa consumption were analysed in this study, including theobromine (TB), caffeine (CAF), theophylline (TP), paraxanthine (PX), 7-methylxanthine (MX) and theanine, an amino acid prevalent in tea.

Dietary intakes in the TwinsUK sample were estimated using a modified version of the European Prospective Investigation into Cancer and Nutrition (EPIC) food frequency questionnaire (FFO). This version incorporates food items from the EPIC Norfolk study [52]. FFQs were excluded if more than 10 food items were unanswered or if the total energy intake, derived from the FFQ and expressed as a ratio of the subject's estimated basal metabolic rate (calculated using the Harris-Benedict equation), fell outside 2 standard deviations of the mean (below 0.52 or above 2.58), as previously described [53].

Whole blood DNA methylation profiles were generated for the same 509 participants in the discovery TwinsUK sample using the Infinium HumanMethylation450 BeadChip (Illumina). Epigenetic data generation and processing has previously been described [54, 55]. Briefly, minfi [56] was used to exclude samples with median methylated/unmethylated ratio <10.5, and ENmix [57] was used for background correction, dye bias correction and quantile normalisation of the data. Methylation beta-values were estimated for signals with detP <0.000001 and nbead > 3. Finally, probes and samples with >5% missingness were excluded, as were any outlier samples identified by Enmix [57]. Polymorphic or cross-reactive probes were removed. Mass spectrometry and DNA methylation data were not always obtained from the same clinical visit and samples were selected at a maximum of 5 years apart in either direction (median = 0.11 years, mean = -0.09years, SD = 1.45 years). Latency between samples was approximately normally distributed with a slight negative skew (Shapiro-Wilk Test, W = 0.96, p = 6.6e-10, Adjusted Fisher-Pearson = -0.284). Blood cell proportions were estimated following the Houseman et al. method [58] and obtained from Horvath's calculator (https://dnamage.clockfoundation.org) [26].

## **Epigenetic clocks**

Epigenetic clocks in this study were estimated using Horvath's 'New Methylation Age Calculator'. The analyses focused on epigenetic age acceleration estimated as the residuals of epigenetic age adjusted for chronological age as estimated in Horvath's calculator. Analyses focused on two epigenetic clocks including GrimAge acceleration (GrimAgeAccel), selected due to its high predictive ability for time to death [27] and previous use in similar work related to diet quality [59]; and a DNA methylation-based estimator of telomere length (DNAmTL) [29]. Additional analyses extended to other epigenetic clocks including the Hannum clock [25], PhenoAge [60], and DunedinPACE [28].

#### Replication cohort data

Replication was undertaken in 1,160 fasting serum samples (median age = 60, median BMI = 27) from the KORA (Cooperative Health Research in the Region of Augsburg) cohort. This was a mixed cohort of males and females (568 and 592 respectively), with 446 neversmokers.

The KORA (Kooperative Gesundheitsforschung in der Region Augsburg) F4 (2006–2008) is a follow-up study from the KORA S4 (n = 4,261) survey carried out 1999–2000 [35]. Fasting blood serum samples were collected from participants of the KORA study

population and profiled using the Metabolon platform (Metabolon, Inc., Durham, NC, USA), as described previously [61]. Median-normalisation was achieved by multiplying each metabolite with overall median values and log-transformed.

Whole blood DNAm profiles in the KORA cohort were generated using the HumanMethylation450 BeadChip, and processing of data has previously been described [62]. Estimation of epigenetic ageing clocks followed the methodology outlined for the discovery sample analysis.

## Statistical analysis

Association analyses were carried out in RStudio (2023.09.1+494) using linear mixed-effects models (R package 'lme4'). Epigenetic acceleration measures were the response variable and theobromine levels were the predictors. Models were adjusted for covariates including blood cell type proportions, age and bodymass index (BMI) as fixed-effect variables, and for family relatedness as a random effect term.

The primary analysis investigated associations between each metabolite and GrimAgeAccel and DNAmTL. Extended analyses also considered additional epigenetic clocks (Hannum clock, PhenoAge, and DunedinPACE).

Sensitivity analyses included additional covariates CAF, TP, PX and MX to account for potential confounding across food components. As multicollinearity is a potential factor in these analyses, two additional sensitivity analyses were undertaken, 10-fold cross validated LASSO and elastic-net regression (R package 'glmnet'). LASSO regression penalises coefficients by shrinkage, reducing the number of variables to control for multicollinearity and the extent of overfitting. Elastic-net regression also uses penalisation to regularise results and reduce the influence of collinear metabolites, by using a composite of LASSO and Ridge regularisation methods to enable best fit. This approach has complementary strengths to LASSO by providing a more stable feature selection, especially in cases of multicollinearity.

#### **Abbreviations**

CAF: Caffeine; DNAm: DNA methylation; MX: 7-methylxanthine; PX: Paraxanthine; TB: Theobromine; TP: Theophylline.

## **AUTHOR CONTRIBUTIONS**

RS and JTB conceived the study. RS and RC carried out the primary analysis, and PRMG contributed to the

analysis. CG, KS, AP, GK, CM, and MW contributed reagents, materials and input on results interpretation. ARM and CD contributed to results interpretation and write-up. RS, RC, and JTB wrote the manuscript. All authors reviewed and approved the manuscript.

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#### **CONFLICTS OF INTEREST**

The authors declare no conflicts of interest related to this study.

## ETHICAL STATEMENT AND CONSENT

Ethical approval for the discovery sample analyses in TwinsUK was granted under TwinsUK BioBank ethics, approved by North West – Liverpool Central Research Ethics Committee (REC reference 19/NW/0187). The KORA study was performed in line with the principles of the Declaration of Helsinki. Study methods were approved by the Ethics Committee of the Bavarian Chamber of Physicians (REC: #06068). Written informed consent was obtained from all research participants prior to taking part in any research activities.

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#### **REFERENCES**

 Monjotin N, Amiot MJ, Fleurentin J, Morel JM, Raynal S. Clinical Evidence of the Benefits of Phytonutrients in Human Healthcare. Nutrients. 2022; 14:1712. <a href="https://doi.org/10.3390/nu14091712">https://doi.org/10.3390/nu14091712</a> PMID:35565680

 Millar CL, Duclos Q, Blesso CN. Effects of Dietary Flavonoids on Reverse Cholesterol Transport, HDL Metabolism, and HDL Function. Adv Nutr. 2017; 8:226–39.

https://doi.org/10.3945/an.116.014050 PMID:28298268

3. Wisnuwardani RW, De Henauw S, Ferrari M, Forsner M, Gottrand F, Huybrechts I, Kafatos AG, Kersting M, Knaze V, Manios Y, Marcos A, Molnár D, Rothwell JA, et al. Total Polyphenol Intake Is Inversely Associated with a Pro/Anti-Inflammatory Biomarker Ratio in European Adolescents of the HELENA Study. J Nutr. 2020; 150:1610–8.

https://doi.org/10.1093/jn/nxaa064 PMID:32221603

- Zumerle S, Sarill M, Saponaro M, Colucci M, Contu L, Lazzarini E, Sartori R, Pezzini C, Rinaldi A, Scanu A, Sgrignani J, Locatelli P, Sabbadin M, et al. Targeting senescence induced by age or chemotherapy with a polyphenol-rich natural extract improves longevity and healthspan in mice. Nat Aging. 2024; 4:1231–48. <a href="https://doi.org/10.1038/s43587-024-00663-7">https://doi.org/10.1038/s43587-024-00663-7</a>
   PMID:38951692
- 5. Akinboye AJ, Kim K, Choi S, Yang I, Lee JG. Alkaloids in

food: a review of toxicity, analytical methods, occurrence and risk assessments. Food Sci Biotechnol. 2023; 32:1133–58.

https://doi.org/10.1007/s10068-023-01295-0 PMID:37362815

 Dhyani P, Quispe C, Sharma E, Bahukhandi A, Sati P, Attri DC, Szopa A, Sharifi-Rad J, Docea AO, Mardare I, Calina D, Cho WC. Anticancer potential of alkaloids: a key emphasis to colchicine, vinblastine, vincristine, vindesine, vinorelbine and vincamine. Cancer Cell Int. 2022; 22:206.

https://doi.org/10.1186/s12935-022-02624-9 PMID:35655306

7. Ajebli M, Khan H, Eddouks M. Natural Alkaloids and Diabetes Mellitus: A Review. Endocr Metab Immune Disord Drug Targets. 2021; 21:111–30.

https://doi.org/10.2174/187153032066620082112 4817

PMID:32955004

 Santiago LÂM, Neto RNM, Ataíde ACS, Fonseca DCS, Soares EFA, de Sá Sousa JC, Mondego-Oliveira R, Ribeiro RM, Cartágenes MSS, Lima-Neto LG, Carvalho RC, de Sousa EM. Flavonoids, alkaloids and saponins: are these plant-derived compounds an alternative to the treatment of rheumatoid arthritis? A literature review. Clin Phytosci. 2021; 7:58.

https://doi.org/10.1186/s40816-021-00291-3

 Omar F, Tareq AM, Alqahtani AM, Dhama K, Sayeed MA, Emran TB, Simal-Gandara J. Plant-Based Indole Alkaloids: A Comprehensive Overview from a Pharmacological Perspective. Molecules. 2021; 26:2297.

https://doi.org/10.3390/molecules26082297 PMID:33921093

 Zhang J, Morris-Natschke SL, Ma D, Shang XF, Yang CJ, Liu YQ, Lee KH. Biologically active indolizidine alkaloids. Med Res Rev. 2021; 41:928–60. https://doi.org/10.1002/med.21747

PMID:33128409

 Ai X, Yu P, Peng L, Luo L, Liu J, Li S, Lai X, Luan F, Meng X. Berberine: A Review of its Pharmacokinetics Properties and Therapeutic Potentials in Diverse Vascular Diseases. Front Pharmacol. 2021; 12:762654.

https://doi.org/10.3389/fphar.2021.762654 PMID:35370628

12. Chieng D, Canovas R, Segan L, Sugumar H, Voskoboinik A, Prabhu S, Ling LH, Lee G, Morton JB, Kaye DM, Kalman JM, Kistler PM. The impact of coffee subtypes on incident cardiovascular disease, arrhythmias, and mortality: long-term outcomes from the UK Biobank. Eur J Prev Cardiol. 2022; 29:2240–9.

https://doi.org/10.1093/eurjpc/zwac189

PMID:36162818

Sesso HD, Manson JE, Aragaki AK, Rist PM, Johnson LG, Friedenberg G, Copeland T, Clar A, Mora S, Moorthy MV, Sarkissian A, Carrick WR, Anderson GL, and COSMOS Research Group. Effect of cocoa flavanol supplementation for the prevention of cardiovascular disease events: the COcoa Supplement and Multivitamin Outcomes Study (COSMOS) randomized clinical trial. Am J Clin Nutr. 2022; 115:1490–500.

https://doi.org/10.1093/ajcn/nqac055 PMID:35294962

Monteiro JP, Alves MG, Oliveira PF, Silva BM. Structure-Bioactivity Relationships of Methylxanthines: Trying to Make Sense of All the Promises and the Drawbacks. Molecules. 2016; 21:974.

https://doi.org/10.3390/molecules21080974 PMID:27472311

 Eren F, Kabaran S. Evaluation of theobromine content and the relationship between cocoa percentages in dark chocolates. FFHD. 2023; 13:520. <a href="https://doi.org/10.31989/ffhd.v13i10.1141">https://doi.org/10.31989/ffhd.v13i10.1141</a>

 Li H, Roxo M, Cheng X, Zhang S, Cheng H, Wink M. Pro-oxidant and lifespan extension effects of caffeine and related methylxanthines in *Caenorhabditis elegans*. Food Chem X. 2019; 1:100005. <a href="https://doi.org/10.1016/j.fochx.2019.100005">https://doi.org/10.1016/j.fochx.2019.100005</a>
 PMID:31432005

Franco R, Oñatibia-Astibia A, Martínez-Pinilla E. Health benefits of methylxanthines in cacao and chocolate. Nutrients. 2013; 5:4159–73.
 https://doi.org/10.3390/nu5104159

 PMID:24145871

18. El Omari N, Bakrim S, Bakha M, Lorenzo JM, Rebezov M, Shariati MA, Aboulaghras S, Balahbib A, Khayrullin M, Bouyahya A. Natural Bioactive Compounds Targeting Epigenetic Pathways in Cancer: A Review on Alkaloids, Terpenoids, Quinones, and Isothiocyanates. Nutrients. 2021; 13:3714.

https://doi.org/10.3390/nu13113714 PMID:34835969

 Crescenti A, Solà R, Valls RM, Caimari A, Del Bas JM, Anguera A, Anglés N, Arola L. Cocoa Consumption Alters the Global DNA Methylation of Peripheral Leukocytes in Humans with Cardiovascular Disease Risk Factors: A Randomized Controlled Trial. PLoS One. 2013; 8:e65744.

https://doi.org/10.1371/journal.pone.0065744 PMID:23840361

20. Karabegović I, Portilla-Fernandez E, Li Y, Ma J, Maas SCE, Sun D, Hu EA, Kühnel B, Zhang Y, Ambatipudi S, Fiorito G, Huang J, Castillo-Fernandez JE, et al. Epigenome-wide association meta-analysis of DNA methylation with coffee and tea consumption. Nat Commun. 2021; 12:2830. https://doi.org/10.1038/s41467-021-22752-6 PMID:33990564

21. Johansson A, Enroth S, Gyllensten U. Continuous Aging of the Human DNA Methylome Throughout the Human Lifespan. PLoS One. 2013; 8:e67378.

https://doi.org/10.1371/journal.pone.0067378 PMID:23826282

22. Seale K, Horvath S, Teschendorff A, Eynon N, Voisin S. Making sense of the ageing methylome. Nat Rev Genet. 2022; 23:585–605.

https://doi.org/10.1038/s41576-022-00477-6 PMID:35501397

 Rakyan VK, Down TA, Maslau S, Andrew T, Yang TP, Beyan H, Whittaker P, McCann OT, Finer S, Valdes AM, Leslie RD, Deloukas P, Spector TD. Human agingassociated DNA hypermethylation occurs preferentially at bivalent chromatin domains. Genome Res. 2010; 20:434–9.

https://doi.org/10.1101/gr.103101.109 PMID:20219945

24. Bell JT, Tsai PC, Yang TP, Pidsley R, Nisbet J, Glass D, Mangino M, Zhai G, Zhang F, Valdes A, Shin SY, Dempster EL, Murray RM, et al, and MuTHER Consortium. Epigenome-wide scans identify differentially methylated regions for age and agerelated phenotypes in a healthy ageing population. PLoS Genet. 2012; 8:e1002629.

https://doi.org/10.1371/journal.pgen.1002629 PMID:22532803

- Hannum G, Guinney J, Zhao L, Zhang L, Hughes G, Sadda S, Klotzle B, Bibikova M, Fan JB, Gao Y, Deconde R, Chen M, Rajapakse I, et al. Genome-wide methylation profiles reveal quantitative views of human aging rates. Mol Cell. 2013; 49:359–67. https://doi.org/10.1016/j.molcel.2012.10.016
   PMID:23177740
- 26. Horvath S. DNA methylation age of human tissues and cell types. Genome Biol. 2013; 14:R115. <a href="https://doi.org/10.1186/gb-2013-14-10-r115">https://doi.org/10.1186/gb-2013-14-10-r115</a> PMID:24138928
- Lu AT, Quach A, Wilson JG, Reiner AP, Aviv A, Raj K, Hou L, Baccarelli AA, Li Y, Stewart JD, Whitsel EA, Assimes TL, Ferrucci L, Horvath S. DNA methylation GrimAge strongly predicts lifespan and healthspan. Aging (Albany NY). 2019; 11:303–27. <a href="https://doi.org/10.18632/aging.101684">https://doi.org/10.18632/aging.101684</a>
   PMID:30669119
- 28. Belsky DW, Caspi A, Corcoran DL, Sugden K, Poulton R, Arseneault L, Baccarelli A, Chamarti K, Gao X, Hannon E, Harrington HL, Houts R, Kothari M, et al. DunedinPACE, a DNA methylation biomarker of the pace of aging. Elife. 2022; 11:e73420.

https://doi.org/10.7554/eLife.73420 PMID:35029144

29. Lu AT, Seeboth A, Tsai PC, Sun D, Quach A, Reiner AP, Kooperberg C, Ferrucci L, Hou L, Baccarelli AA, Li Y, Harris SE, Corley J, et al. DNA methylation-based estimator of telomere length. Aging (Albany NY). 2019; 11:5895–923. https://doi.org/10.18632/aging.102173

https://doi.org/10.18632/aging.102173 PMID:31422385

Bordoni L, Agostinho de Sousa J, Zhuo J, von Meyenn
 Evaluating the connection between diet quality,
 EpiNutrient intake and epigenetic age: an observational study. Am J Clin Nutr. 2024; 120:1143–55

https://doi.org/10.1016/j.ajcnut.2024.08.033 PMID:39510725

31. Fitzgerald KN, Hodges R, Hanes D, Stack E, Cheishvili D, Szyf M, Henkel J, Twedt MW, Giannopoulou D, Herdell J, Logan S, Bradley R. Potential reversal of epigenetic age using a diet and lifestyle intervention: a pilot randomized clinical trial. Aging (Albany NY). 2021; 13:9419–32.

https://doi.org/10.18632/aging.202913 PMID:33844651

- Waziry R, Ryan CP, Corcoran DL, Huffman KM, Kobor MS, Kothari M, Graf GH, Kraus VB, Kraus WE, Lin DTS, Pieper CF, Ramaker ME, Bhapkar M, et al. Effect of long-term caloric restriction on DNA methylation measures of biological aging in healthy adults from the CALERIE trial. Nat Aging. 2023; 3:248–57. <a href="https://doi.org/10.1038/s43587-022-00357-y">https://doi.org/10.1038/s43587-022-00357-y</a>
   PMID:37118425
- Purkiewicz A, Pietrzak-Fiećko R, Sörgel F, Kinzig M. Caffeine, Paraxanthine, Theophylline, and Theobromine Content in Human Milk. Nutrients. 2022; 14:2196.
   <a href="https://doi.org/10.3390/nu14112196">https://doi.org/10.3390/nu14112196</a>
   PMID:35683994
- Pallister T, Jennings A, Mohney RP, Yarand D, Mangino M, Cassidy A, MacGregor A, Spector TD, Menni C. Characterizing Blood Metabolomics Profiles Associated with Self-Reported Food Intakes in Female Twins. PLoS One. 2016; 11:e0158568.
   <a href="https://doi.org/10.1371/journal.pone.0158568">https://doi.org/10.1371/journal.pone.0158568</a>
   PMID:27355821
- 35. Holle R, Happich M, Löwel H, Wichmann HE, and MONICA/KORA Study Group. KORA--a research platform for population based health research. Gesundheitswesen. 2005 (Suppl 1); 67:S19–25. https://doi.org/10.1055/s-2005-858235
  PMID:16032513
- 36. Hukkanen J, Jacob P 3rd, Peng M, Dempsey D,

Benowitz NL. Effect of nicotine on cytochrome P450 1A2 activity. Br J Clin Pharmacol. 2011; 72:836–8. https://doi.org/10.1111/j.1365-2125.2011.04023.x PMID:21599724

37. Marioni RE, Harris SE, Shah S, McRae AF, von Zglinicki T, Martin-Ruiz C, Wray NR, Visscher PM, Deary IJ. The epigenetic clock and telomere length are independently associated with chronological age and mortality. Int J Epidemiol. 2018; 45:424–32. <a href="https://doi.org/10.1093/ije/dyw041">https://doi.org/10.1093/ije/dyw041</a>
PMID:27075770

38. Kabacik S, Lowe D, Fransen L, Leonard M, Ang SL, Whiteman C, Corsi S, Cohen H, Felton S, Bali R, Horvath S, Raj K. The relationship between epigenetic age and the hallmarks of aging in human cells. Nat Aging. 2022; 2:484–93.

https://doi.org/10.1038/s43587-022-00220-0 PMID:37034474

39. Thorn CF, Aklillu E, McDonagh EM, Klein TE, Altman RB. PharmGKB summary: caffeine pathway. Pharmacogenet Genomics. 2012; 22:389–95. https://doi.org/10.1097/FPC.0b013e3283505d5e PMID:22293536

 Grzegorzewski J, Bartsch F, Köller A, König M. Pharmacokinetics of Caffeine: A Systematic Analysis of Reported Data for Application in Metabolic Phenotyping and Liver Function Testing. Front Pharmacol. 2022; 12:752826.

https://doi.org/10.3389/fphar.2021.752826 PMID:35280254

Baggott MJ, Childs E, Hart AB, de Bruin E, Palmer AA, Wilkinson JE, de Wit H. Psychopharmacology of theobromine in healthy volunteers. Psychopharmacology (Berl). 2013; 228:109–18. <a href="https://doi.org/10.1007/s00213-013-3021-0">https://doi.org/10.1007/s00213-013-3021-0</a>
 PMID:23420115

 Yoneda M, Sugimoto N, Katakura M, Matsuzaki K, Tanigami H, Yachie A, Ohno-Shosaku T, Shido O. Theobromine up-regulates cerebral brain-derived neurotrophic factor and facilitates motor learning in mice. J Nutr Biochem. 2017; 39:110–6. <a href="https://doi.org/10.1016/j.jnutbio.2016.10.002">https://doi.org/10.1016/j.jnutbio.2016.10.002</a>
 PMID:27833051

 Hahn J, Virk HUH, Wang Z, Tang WHW, Krittanawong C. Abstract 14006: Cardiovascular Health Benefits of Theobromine in Cacao and Chocolate. Circulation. 2021 (Suppl 1); 144:A14006. https://doi.org/10.1161/circ.144.suppl 1.14006

44. Abdel-Hady H, Nasef N, Shabaan AE, Nour I. Caffeine therapy in preterm infants. World J Clin Pediatr. 2015; 4:81–93.

https://doi.org/10.5409/wjcp.v4.i4.81 PMID:26566480  Martínez-Pinilla E, Oñatibia-Astibia A, Franco R. The relevance of theobromine for the beneficial effects of cocoa consumption. Front Pharmacol. 2015; 6:30. https://doi.org/10.3389/fphar.2015.00030 PMID:25750625

46. Peron G, Gargari G, Meroño T, Miñarro A, Lozano EV, Escuder PC, González-Domínguez R, Hidalgo-Liberona N, Del Bo' C, Bernardi S, Kroon PA, Carrieri B, Cherubini A, et al. Crosstalk among intestinal barrier, gut microbiota and serum metabolome after a polyphenol-rich diet in older subjects with "leaky gut": The MaPLE trial. Clin Nutr. 2021; 40:5288–97. <a href="https://doi.org/10.1016/j.clnu.2021.08.027">https://doi.org/10.1016/j.clnu.2021.08.027</a>
PMID:34534897

 McCartney DL, Stevenson AJ, Hillary RF, Walker RM, Bermingham ML, Morris SW, Clarke TK, Campbell A, Murray AD, Whalley HC, Porteous DJ, Visscher PM, McIntosh AM, et al. Epigenetic signatures of starting and stopping smoking. EBioMedicine. 2018; 37:214– 20.

https://doi.org/10.1016/j.ebiom.2018.10.051 PMID:30389506

48. Sansone R, Ottaviani JI, Rodriguez-Mateos A, Heinen Y, Noske D, Spencer JP, Crozier A, Merx MW, Kelm M, Schroeter H, Heiss C. Methylxanthines enhance the effects of cocoa flavanols on cardiovascular function: randomized, double-masked controlled studies. Am J Clin Nutr. 2017; 105:352–60.

https://doi.org/10.3945/ajcn.116.140046 PMID:28003203

49. Raman G, Avendano EE, Chen S, Wang J, Matson J, Gayer B, Novotny JA, Cassidy A. Dietary intakes of flavan-3-ols and cardiometabolic health: systematic review and meta-analysis of randomized trials and prospective cohort studies. Am J Clin Nutr. 2019; 110:1067–78.

https://doi.org/10.1093/ajcn/nqz178 PMID:31504087

 Verdi S, Abbasian G, Bowyer RCE, Lachance G, Yarand D, Christofidou P, Mangino M, Menni C, Bell JT, Falchi M, Small KS, Williams FMK, Hammond CJ, et al. TwinsUK: The UK Adult Twin Registry Update. Twin Res Hum Genet. 2019; 22:523–9.

https://doi.org/10.1017/thg.2019.65 PMID:31526404

51. Long T, Hicks M, Yu HC, Biggs WH, Kirkness EF, Menni C, Zierer J, Small KS, Mangino M, Messier H, Brewerton S, Turpaz Y, Perkins BA, et al. Wholegenome sequencing identifies common-to-rare variants associated with human blood metabolites. Nat Genet. 2017; 49:568–78.

https://doi.org/10.1038/ng.3809

PMID:28263315

 Day N, Oakes S, Luben R, Khaw KT, Bingham S, Welch A, Wareham N. EPIC-Norfolk: study design and characteristics of the cohort. European Prospective Investigation of Cancer. Br J Cancer. 1999 (Suppl 1); 80:95–103.

PMID: 10466767

53. Teucher B, Skinner J, Skidmore PM, Cassidy A, Fairweather-Tait SJ, Hooper L, Roe MA, Foxall R, Oyston SL, Cherkas LF, Perks UC, Spector TD, MacGregor AJ. Dietary patterns and heritability of food choice in a UK female twin cohort. Twin Res Hum Genet. 2007; 10:734–48.

https://doi.org/10.1375/twin.10.5.734 PMID:17903115

- 54. Kurushima Y, Tsai PC, Castillo-Fernandez J, Couto Alves A, El-Sayed Moustafa JS, Le Roy C, Spector TD, Ide M, Hughes FJ, Small KS, Steves CJ, Bell JT. Epigenetic findings in periodontitis in UK twins: a cross-sectional study. Clin Epigenetics. 2019; 11:27. https://doi.org/10.1186/s13148-019-0614-4 PMID:30760334
- 55. Costeira R, Evangelista L, Wilson R, Yan X, Hellbach F, Sinke L, Christiansen C, Villicaña S, Masachs OM, Tsai PC, Mangino M, Menni C, Berry SE, et al. Metabolomic biomarkers of habitual B vitamin intakes unveil novel differentially methylated positions in the human epigenome. Clin Epigenetics. 2023; 15:166.

https://doi.org/10.1186/s13148-023-01578-7 PMID:37858220

56. Aryee MJ, Jaffe AE, Corrada-Bravo H, Ladd-Acosta C, Feinberg AP, Hansen KD, Irizarry RA. Minfi: a flexible and comprehensive Bioconductor package for the analysis of Infinium DNA methylation microarrays. Bioinformatics. 2014; 30:1363–9.

https://doi.org/10.1093/bioinformatics/btu049 PMID:24478339

57. Xu Z, Niu L, Li L, Taylor JA. ENmix: a novel background correction method for Illumina HumanMethylation450 BeadChip. Nucleic Acids Res. 2016; 44:e20.

https://doi.org/10.1093/nar/gkv907 PMID:26384415  Houseman EA, Accomando WP, Koestler DC, Christensen BC, Marsit CJ, Nelson HH, Wiencke JK, Kelsey KT. DNA methylation arrays as surrogate measures of cell mixture distribution. BMC Bioinformatics. 2012; 13:86. https://doi.org/10.1186/1471-2105-13-86

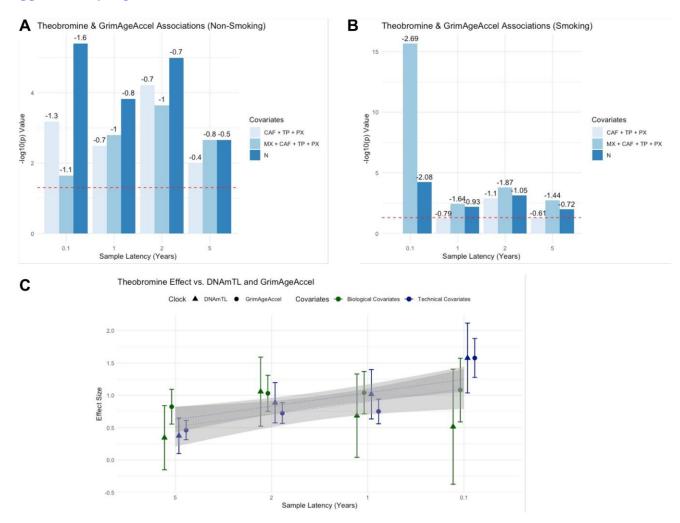
https://doi.org/10.1186/1471-2105-13-86 PMID:22568884

- Reynolds LM, Houston DK, Skiba MB, Whitsel EA, Stewart JD, Li Y, Zannas AS, Assimes TL, Horvath S, Bhatti P, Baccarelli AA, Tooze JA, Vitolins MZ. Diet Quality and Epigenetic Aging in the Women's Health Initiative. J Acad Nutr Diet. 2024; 124:1419–30.e3. <a href="https://doi.org/10.1016/j.jand.2024.01.002">https://doi.org/10.1016/j.jand.2024.01.002</a>
   PMID:38215906
- Levine ME, Lu AT, Quach A, Chen BH, Assimes TL, Bandinelli S, Hou L, Baccarelli AA, Stewart JD, Li Y, Whitsel EA, Wilson JG, Reiner AP, et al. An epigenetic biomarker of aging for lifespan and healthspan. Aging (Albany NY). 2018; 10:573–91. <a href="https://doi.org/10.18632/aging.101414">https://doi.org/10.18632/aging.101414</a>
   PMID:29676998
- Pallister T, Haller T, Thorand B, Altmaier E, Cassidy A, Martin T, Jennings A, Mohney RP, Gieger C, MacGregor A, Kastenmüller G, Metspalu A, Spector TD, Menni C. Metabolites of milk intake: a metabolomic approach in UK twins with findings replicated in two European cohorts. Eur J Nutr. 2017; 56:2379–91. https://doi.org/10.1007/s00394-016-1278-x PMID:27469612
- 62. Gomez-Alonso MDC, Kretschmer A, Wilson R, Pfeiffer L, Karhunen V, Seppälä I, Zhang W, Mittelstraß K, Wahl S, Matias-Garcia PR, Prokisch H, Horn S, Meitinger T, et al. DNA methylation and lipid metabolism: an EWAS of 226 metabolic measures. Clin Epigenetics. 2021; 13:7.

https://doi.org/10.1186/s13148-020-00957-8 PMID:33413638

## **SUPPLEMENTARY MATERIALS**

## **Supplementary Figure**



Supplementary Figure 1. Sensitivity analyses exploring the specificity of the theobromine association with epigenetic ageing measures in the TwinsUK sample. (A, B) Theobromine and GrimAge association significance in never smokers and current/ex-smokers. Covariates in the models included 'N' (BMI + Age + Cell Proportions), as well as 'N' and methylxathines CAF, TP, PX and MX, as labelled on the Figure. TB coefficient effect sizes are shown. (C) Magnitude of effect sizes of DNAmTL association with GrimAgeAccel. As the scales and direction differ (with negative GrimAgeAccel and positive DNAmTL both indicating a slower age) the regression coefficient values of GrimAgeAccel were converted to absolute values and maximum values of DNAmTL were scaled to GrimAgeAccel (scale factor: 43.43). Correlation lines between latency values are applied (linear model) and standard errors are shown.

## **Supplementary Table**

Please browse Full Text version to see the data of Supplementary Table 1.

## Supplementary Table 1. Association results in TwinsUK cohort.

Sample Latency is provided in years between methylation and metabolomic measurements. Covariates included in the linear model are 'N' (BMI + Age + Cell Proportions) and methylxathines CAF, TP, PX and MX as labelled.